

**Detection and Management of Early Blight of Potato  
Caused by *Alternaria alternata***

अल्टरनेरिया अल्टरनेटा द्वारा जनित आलू की अगेती अगंमारी रोग का  
अनुसन्धान एवं प्रबंधन

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THESIS

**MASTER OF SCIENCE IN AGRICULTURE  
(PLANT PATHOLOGY)**



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Agriculture University, Kota**

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Thesis

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**By**

**Priyanka Kumari Meena**

**2020**

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## Chapter 1

### INTRODUCTION

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The potato (*Solanum tuberosum*) is an important food crop in world as well as in India and important vegetable crops, belonging to family solanaceae. Potato originated from the hills of Andes and Bolivia in South America. It was introduced in Europe by Spaniards in the second half of sixteenth century; from there it spreads throughout Europe and rest of the world in the mid-seventeenth to mid-eighteenth century. It was introduced in India by *Portuguese* in the seventeenth century (Chakraborty, 2012). Potato is grown in more than 100 countries, under temperate, subtropical and tropical conditions. It is essentially a "cool weather crop", with temperature being the main limiting factor on production of tuber growth is sharply inhibited in temperatures below 10°C (50°F) and above 30°C (86°F), while optimum yields are obtained where mean daily temperatures are in the 18 to 20°C (64 to 68°F) range (FAO, 2008). India ranks as second world's largest potato producing nation after China. The major potato producing states are Uttar Pradesh, West Bengal, Bihar, Gujarat, Madhya Pradesh, Punjab, Haryana, Assam, Jharkhand and Chhattisgarh.

In Rajasthan total area of vegetables 168.55 ha and production of 1767.53 MT. In potato 13819 ha and production 279.54 MT and productivity 201.6 q/ha. The major potato producing districts are Dholpur, Bharatpur, Hanumangarh, Kota, Sirohi, Srigangangar and Jalore (State Department of Horticulture and Agriculture, 2018-19). In India total area of vegetables 10436 ha and production 187474 MT. (NHB 2018-19). Whereas total area in potato 2151000 ha and production 48529000 MT and productivity 225611 kg/ha (FAO 2018-19). The potato can be grown almost in any type of soil, except saline and alkaline soils. Naturally loose soils which offer the least resistance to enlargement of the tubers are preferred loamy and sandy loam soils that are rich in organic matter with good drainage and aeration are the most suitable. Soil with a pH range of 5.2-6.4 is considered ideal for its cultivation (FAO, 2008). Potato is a valuable source of carbohydrate and can be used both for table consumption as well as processed products. Fresh potato tubers contain around 80% water and 20% dry matter. More than 75% of dry matter is starch but it also contains protein, fibre and small amount of fatty acids (Prokop and Albert, 2008). It is also rich in minerals such as potassium, phosphorus, magnesium and vitamins like B<sub>1</sub>, B<sub>3</sub> and B<sub>6</sub> (Camire *et al.*, 2009). It is one of mankind's most valuable food crops (FAO,

2004). Potatoes are rich in protein, calcium and vitamin C and have an especially good amino acid balance. A single medium-sized potato contains about half the daily adult requirement of vitamin C other staples such as rice and wheat have none. Boiled potato has more protein than maize, and nearly twice the calcium (Umadevi *et al.*, 2013). The intensive and extensive cultivation under the most favorable environmental conditions for potato crop production in the state failed to provide significant strides in potato yields, because of a number of production constraints, of which may be due to frequent occurrence of fungi, bacteria and viruses (Khurana, 2004). Losses can occur when crops are growing, at lifting and when tubers are stored. Some diseases do not destroy tubers, but the surface blemishes they cause decrease marketable value. It is therefore, much emphasis should be given on the production and use of high quality and disease free seed. Though the list of pathogens infecting potato has remained almost unchanged over many decades, there has been steady expansion in the area under potato and improvement in tuber yield/productivity (CIP, 1996). Disease control is a prerequisite for improving and maintaining yield and quality of the potato crop and since the potato became widely grown serious outbreaks of disease and crop failures and consequent social and economic effects have repeatedly provided incentive for improvement. Early blight is one of the most important foliar diseases of potato (Christ, 1989; Pelletier and Fry, 1989; Steinberg *et al.*, 1989 and Vanderwaals *et al.*, 2001). It is caused by two species of genus *Alternaria*, i.e. *Alternaria solani* and *Alternaria alternata* worldwide on potato crops (Gudmestad and Pasche, 2007). *Alternaria solani* and *Alternaria alternata* both are more risk important pathogens on potato crops and significant impact on the tuber yield (Hausladen and Leiminger, 2007 and Kapsa, 2007). The symptoms on the foliage can appear at any stage of the growth and causes characteristic leaf spots and blight. Normally the disease symptoms become apparent during tuber bulking stage and develop leading to the harvest. The early blight is first observed on the plants as small, black lesions mostly on the older foliage. Spots enlarge, and by the time they are one-fourth inch in diameter or larger and concentric rings in a bull's eye pattern can be seen in the center of the diseased area. Tissue surrounding the spots may turn yellow. If high temperature and humidity occur at this time, much of the foliage is killed. Lesions on the stems are similar to those on leaves, sometimes girdling the plant if they occur near the soil line (Kemmitt, 2002). Due to this disease, heavy losses on crop productivity and tuber quality recorded. 5 to 40% of yield losses have

been reported in Israel (Rotem and Feldman, 1965) and 20 to 30% in the USA (Christ and Maczuga, 1989; Shtienberg *et al.*, 1989). The average annual yield losses due to early blight vary enormously from 5-78% but it depends on weather conditions and type of variety grown (Waals *et al.*, 2004 and Pasche *et al.*, 2004, 2005). Young plants are relatively resistant, but the susceptibility increase gradually and continuously from the initiation of tuber formation so that mature plants are most susceptible to the disease (Campo *et al.*, 2001). Reducing both the quantity and quality of marketable tubers (Nnodu *et al.*, 1982). Environmental factors such as temperature, wetness duration and relative humidity affect the development of early blight on potatoes (Harrison *et al.*, 1965; Adams and Stevenson, 1990; Vloutoglou and Kalogerakis, 2000). Temperature increases *Alternaria solani* infection and sporulation (Vloutoglou and Kalogerakis, 2000). Water in the form of high relative humidity, rainfall or dew accumulation can increase conidia germination and pathogen infection (Rotem, 2004). Alternating low and high humidity conditions have also been shown to favour disease development (Van der Waals *et al.*, 2001). Potato is a source of both food and income in the growing countries of the world which able to change greatly the food security of countries because of its high productivity per unit area and time compared to other crops. The potato chips and wafers are popular processed food items that give considerable value addition to potato. Incidence of early blight has been observed in considerable form in potato growing areas of South Eastern Rajasthan during *Rabi* season. In view of the seriousness of the disease, the studies were planned to undertake on the following aspects which could help in the management of the disease.

- i. To study the symptoms, isolation, purification, pathogenicity and identification of pathogen.
- ii. To study the efficacy of bio-agents and media against the pathogen *in vitro* condition.
- iii. Screening the genotypes against the disease.
- iv. To study the bio-efficacy of fungicide against the pathogen *in vitro* and *in vivo* condition.



## Chapter 2

### REVIEW OF LITERATURE

---

#### 2.1 To study the symptoms, isolation, purification, identification and pathogenicity of pathogen

Ellis (1971) described symptoms round, oval or irregular, brown or dark brown, often concentrically ridged target spots and under microscopy observed solidity and beaked conidia 9 to 11 transverse septa and a few or no longitudinal or oblique septa.

Pathak (1972) the culture of pathogen was purified by hyphal tip method.

Johnston and Booth (1983) studied isolation of pathogen with segments of diseased tissue along with some healthy leaf portion (5×5 mm<sup>2</sup>) were cut with a sterilized razor blade at the margins of the diseased spots on the leaves and surface sterilized in 0.1% mercuric chloride solution for 30 second and single spore technique was used for purification. The leaf symptoms are characteristic dark brown reddish, yellow or mainly and greenish yellow in colour.

Foolad *et al.* (2008) reported that symptoms of disease small, dark and circular lesions flatter distinctly zonate as they expand and stem lesions occur as roughly circular, sunken, dark.

Kumar *et al.* (2008) found that isolation of *Alternaria* spp. showed pigmentation varied from yellow, brown, black, brownish to green black on potato dextrose agar medium.

Schultz and French (2009) observed that infected stems show sunken, elongated spots that may also show the typical concentric rings and lesions in tubers appear as slightly sunken dark irregular spots with raised borders.

Verma and Verma (2010) described the genus is characterized by the formation of polymorphous conidia either singly or in short or longer chains and provided with cross, longitudinal, as well as oblique septa and having longer or short beaks.

Roopa (2012) tested pathogenicity by inoculating with spore suspension and homogenized mycelial bits ( $2 \times 10^4$  spores/ml) of *Alternaria solani* on foliage of 30 days old and proved the Koch's postulates.

Ganie *et al.* (2013) proved the pathogenicity by using spore suspension from 15 days old culture with concentration of  $2 \times 10^4$  conidia/ml on Kufri Jyoti variety of

potato and revealed that the initiation of typical symptoms of the disease appeared after 10 days of inoculation on injured detached leaves of potato. They also revealed that symptoms in early stages of disease development, small irregular to circular dark brown spots on lower leaves appear, measuring 0.5 mm in size. After 4 weeks concentric rings form as a result of irregular growth patterns by the organism in the leaf tissue giving the lesion a characteristic target spot or bull eye appearance.

Mamgain *et al.* (2013) reported that spore was attached in chains, multi-celled and pigmented. The symptoms appear as small, circular or irregular, dark- brown to black spots on the older (lower) leaves. These spots enlarge up to  $\frac{3}{8}$  inch in diameter and gradually may become angular-shaped. Initial lesions on young, fully expanded leaves may confuse with brown spot lesions. These first lesions appear about two to three days after infection observed by Bauske (2018).

Robinson (2018) reported that chlorotic symptoms may develop on infected leaves through time as lesions coalesce and clusters of infection form. Elongated, brown to black lesions may develop on the stems and petioles of infected plants. The infected tubers appear as dark-coloured, sunken lesions on the tuber surface and tuber lesions may be circular or irregular in shape and often are accompanied by a large, raised dark-brown border.

## **2.2 To study the efficacy of bio-agent and media against the pathogen *in vitro* condition**

### **2.2.1 Efficacy of media study**

Arunkumar (2006) recorded maximum growth of *Alternaria solani* after nine days of incubation in potato dextrose broth.

Roopa (2012) found maximum dry mycelia weight (249.27 mg) of *Alternaria solani* after 9<sup>th</sup> day of inoculation in potato dextrose broth.

Somappa *et al.* (2013) observed maximum radial growth on Czapeck's dox agar medium (50 mm) followed by Potato dextrose agar (37 mm).

Koley and Mahapatra (2015) tested the growth of pathogen in twelve liquid and solid media and revealed that potato dextrose agar and oat meal agar among solid media and Richard's broth and Sabouraud's broth among liquid media seemed to be better for growth of *Alternaria solani*. Dry weight of *Alternaria solani* was maximum in potato dextrose broth medium (289.33 mg) followed by oat meal broth medium (281.33 mg) and Czapek's Dox broth (240.67 mg). The colour of the colony

*Alternaria solani* was having dark brown in case of potato dextrose agar medium, Oat meal agar medium imparted grey colour of the colony, Czapek's agar medium whitish colony with greenish border and substrate light colour. Maximum sporulation was observed on oat meal agar followed by potato dextrose agar medium. However, moderate sporulation was observed on Czapek's dox agar.

Kumar *et al.* (2018c) recorded highest radial growth of *Alternaria solani* on potato dextrose agar (63.00 mm) followed by oat meal agar (42.33 mm) and Czapek's dox agar (27.73 mm).

### 2.2.2 Efficacy of bio-agent

Yadav and Pathak (2011) observed that *Trichoderma viride* @ 0.5% was efficacious in controlling early blight of potato.

Sharma *et al.* (2014) reported that *Trichoderma harzianum* and *Trichoderma viride* are the widely used species and have been exploited on about 87 different crops and about 70 soils borne as well as 18 foliar pathogens, respectively. *Trichoderma* as a potential bio-control agent against *Alternaria* observed by Kushwaha and Verma (2014).

Murmu *et al.* (2015) found *Trichoderma viride* highly effective in percent disease reduction (52.39%) against early blight of potato with highest tuber yield (25.51 t/ha) as compared to control (19.53 t/ha).

Mahantesh *et al.* (2017) reported that *Trichoderma harzianum* showed the maximum percent inhibition (80.36%) followed by *Trichoderma viride* (75.86%) against *Alternaria solani* at UAHS, shivamogga.

Yadav *et al.* (2018) observed that *Trichoderma harzianum* at 3000 ppm concentration significantly inhibit the radial growth (4.88 mm) of *Alternaria solani* as compare to control (15.03 mm) *in vitro* condition.

Kumar *et al.* (2018b) recorded that maximum percent inhibition (78.97%) of the *Alternaria solani* by *Trichoderma viride* followed by *Trichoderma harzianum* inhibition percent (75.33%) as compare to control (87.16 mm).

Sarfraz *et al.* (2018) observed that *Trichoderma harzianum* at (48, 96, 144 and 172 hrs) growth inhibition (7.50, 12.90, 29.44 and 43.00%) followed by *Trichoderma viride* growth inhibition (4.40, 9.10, 29.10 and 37.29%) against *Alternaria solani* causing by early blight of potato.

Sreenivasunu *et al.* (2019) recorded that the radial growth (32.5 mm) and percent inhibition (47.29%) of *Alternaria solani* by using *Trichoderma viride*.

### 2.3 Screening the genotypes against the disease

Dey and Chakraborty (2012) evaluated ten varieties like K. Chandramukhi, K. Pukhraj, K. Chipsona-1, K. Jawahar, K. Anand, K. Giriraj, K. Surya and K. Bahar and maximum incidence and intensity percent were observed in the variety K. Giriraj (43.68 and 27.73%) followed by K. Anand (40.45 and 25.87%), K. Jawahar (33.67 and 23.87%), K. Pukhraj (27.08 and 22.27%), and K. Surya (18.95 and 6.27%) and K. Chipsona - 1 (15.31 and 7.33%) 21 days after first appearance of the symptoms at Adisaptagram block seed farm.

Mehboob *et al.* (2013) screened 29 test lines against early blight of potato. 15 lines including cardinal and desire were found to be very highly susceptible. 8 lines including diamante FST-white and PPS 9813 showed highly susceptible response. 2 lines that are 9802 and 396266-33 were susceptible. FD-18 was found to be resistant, while two lines such as F3-39 and FD-48-41 were so shown moderately resistant response.

Ganie and Ghani (2013) screened twenty five potato genotypes for early blight of potato under natural conditions. Out of twenty five, one genotype SM/92-338 showed tolerant reaction while three genotypes *viz.*, Kufri Himan, SM/96-27 and SM/94-44 were moderately tolerant. Nine genotypes *viz.*, Kufri Girdari, Kufri Shailaja, Kufri Chandramukhi, SM/98-239, SM/93-237, SM/90-45, HB/82-18, HB/50-45 and Shalimar potato-1 were moderately susceptible.

Kumar *et al.* (2015) evaluated genotypes against early blight of potato at main experiment station (MES) of Vegetable Science in Narendra Deva University of Agriculture and Technology, Kumarganj, Faizabad, Uttar Pradesh, Out of them, Kufri Pukhraj and Kufri Pushkar were found moderately resistant, remaining K. Bahar and Sindhuri found susceptible and K. Sutlej found highly susceptible.

Prabha and Nanda (2017) were screened 18 clonal bulk population, 43 F<sub>1</sub>C<sub>2</sub> and 90 F<sub>1</sub>C<sub>3</sub> clonal progenies of potato for resistance against *Alternaria solani* under natural and artificial conditions. All the genotypes of clonal bulks, F<sub>1</sub>C<sub>2</sub> and F<sub>1</sub>C<sub>3</sub> progenies showed moderate resistance reactions against blight incidence and most of the genotypes of clonal bulks, F<sub>1</sub>C<sub>2</sub> and F<sub>1</sub>C<sub>3</sub> clonal progenies showed moderate resistance against blight severity. Among artificial inoculated genotypes of clonal bulks and progenies, the genotype CIP 398201 of clonal bulk, four genotypes

(CIP 302024-11-2, CIP 398201-19-1, CIP 398201-19-2 and CIP 398201-20-1) of F<sub>1</sub>C<sub>2</sub> progenies and five genotypes (CIP 398201-7-2-1, CIP 398201-15-4-1, CIP 398201-5-3-1, CIP 398201-11-6-1 and CIP 398201-2-2-1) of F<sub>1</sub>C<sub>3</sub> progenies were found moderately resistant against blight incidence, while remaining genotypes showed moderately susceptible reaction.

#### **2.4 To study the bio-efficacy of fungicide against the pathogen *in vitro* and *in vivo* condition**

Arunkumar (2006) recorded maximum inhibition of Propiconazole (0.1%), Metalaxyl MZ (0.2%) and Perfekt (0.3%) against *Alternaria solani* under *in vitro* condition, while Propiconazole (0.1%), Pyraclostrobin (0.2%) were found to be most effective in reducing the severity of the disease and increasing yield over control under field conditions.

Singh and Singh (2006) tested the efficacy of seven fungicides *viz.* Chlorothalonil, Copper Oxychloride, Azoxystrobin, Propineb, Copper hydroxide, Mancozeb at 2500, 2000, 1000, 500 and 250 ppm and Hexaconazole at 1000, 500, 200, 100 and 50 ppm against *Alternaria solani*, respectively and observed that all the fungicides significantly reduced the radial growth of the *Alternaria solani*. However, Hexaconazole was give 100% growth inhibition.

Kumar *et al.* (2007) recorded that Hexaconazole (0.05%) and Azoxystrobin (0.2%) was very effective in controlling early blight. Pyraclostrobin significantly reduced the early blight and increased the yield in tomato and potato reported by Ganesham and Chethana (2009).

Issiakhem and Bouznad (2010) tested Difenconazole and Chlorothalonil on conidial germination and mycelial growth of *Alternaria solani* under *in vitro* conditions. The results revealed that Difenconazole had a better effectiveness than Chlorothalonil in inhibition of mycelial growth and conidial germination.

Horsfield *et al.* (2010) evaluated protective and curative activity of fungicides in glasshouse and field for the control of early blight. Boscalid, Azoxystrobin and Difenconazole were highly effective in the control of early blight when applied up to three days before or three days after inoculation.

Jambhulkar *et al.* (2012) reported that Azoxystrobin 23% SC reducing early blight disease 38.9% as compare to control in tomato crop.

Genie (2012) evaluated different systematic and non-systematic fungicide against early blight of potato crop and reported that Mancozeb (17.76%) and

Hexaconazole (14.37%) found effective in reducing disease severity as compare to control (34.38%).

Sahu *et al.* (2013) reported that Mancozeb 75% WP (42.36%) and Pyraclostrobin 38% WG (35.73%) significantly reduced percent disease intensity with increased the yield (29.75 and 31.81 t/ha) as compared to control (60.80%) and yield (21.15 t/ha) in early blight disease in tomato.

Herle *et al.* (2014) reported that different fungicides tested against mycelial growth of *Alternaria solani* were Mancozeb 75% WP at (0.1, 0.2 and 0.25%) concentrations, percent inhibition (57.22, 65.23 and 99.33%) and Hexaconazole 5% EC and Propiconazole 25% EC at (0.05, 0.1 and 0.15%) concentrations are equally effective and significantly superior with 100% inhibition, which are on par with Difenconazole 25% EC (100% inhibition) at 0.1 and 0.25% and (91.11%) at 0.05% concentration in laboratory condition. Under field conditions, the minimum percent disease incidence by using different fungicides such as Mancozeb 75% WP at (35, 50, 65, 75 DAS) percent inhibition (14.13, 32.00, 40.67 and 52.00%), Difenconazole 25% EC (12.30, 21.00, 25.67 and 33.16%), Hexaconazole 5% EC (11.96, 18.50, 20.33 and 28.66%) and Propiconazole 25% EC (13.66, 16.00, 25.00 and 31.83%) maximum percent disease incidence was noticed in Mancozeb 75% WP (52%) at 72 days after sowing.

Stepanovic *et al.* (2015) observed that the tested *Alternaria solani* isolates were also highly sensitive to Difenconazole. All isolates were able to grow well at 0.001ug/ml Difenconazole concentration (<10% growth inhibition), but they were partially inhibited by the next two higher concentrations (0.01 and 0.1 ug/ml) and significantly inhibited by the concentration of 10 ug/ml (>95% growth inhibition). The values for Difenconazole ranged between 0.018 and 0.037 ug/ml. The growth inhibition of over 82% was achieved by Pyraclostrobin concentration of 10 ug/ml. Concentrations which inhibited mycelia growth by 50% (EC<sub>50</sub>) ranged from 0.0014 to 0.0041 ug/ml.

More *et al.* (2016) conducted field experiments on efficacy of sequential sprays of different fungicides against early blight variety Kufri Pukhraj in potato for consecutive three years and revealed that spray of Hexaconazole 5 EC (0.05%) @ 0.5 ml/litre of water lower the disease intensity at 10 days after first spraying (10.33%), 20 DAS (13.05%) and 30 DAS (16.27%) with tuber yield (15.00 t/ha) as

compare to control 10 DAS (16.23%), 20 DAS (25.20%) and 30 DAS (30.87%) with tuber yield (11.96 t/ha).

More *et al.* (2016) observed minimum disease incidence and intensity of early blight Mancozeb 75% WP (20.83 and 16.16.26%) with increased the yield (16.13 t/ha) and Hexaconazole 5% EC (21.63 and 16.27%) with increased the yield (15.00 t/ha) as compared to control (45.06 and 30.87%) yield (11.96 t/ha) in potato crop (at 10 days after first spray (14.54%), 20 DAS (17.27%) and 30 DAS (21.63%) in potato crop.

Mahantesh *et al.* (2017) observed that Propiconazole @ 100, 250, 500, 1000 ppm showed 80.08, 83.48, 86.42 and 90.58% inhibition followed by Hexaconazole @ 100, 250, 500, and 1000 ppm and inhibition 80.75, 83.18, 85.12, 87.80% and then Difenconazole @ 100, 250, 500, and 1000 ppm with 78.24, 81.20, 84.63 and 88.07% inhibition and Azoxystrobin @ 100, 250, 500 and 1000 ppm with 75.36, 78.39, 81.86 and 83.53% inhibition and then Mancozeb 75 WP @ 100, 250, 500 and 1000 ppm and per cent inhibition of 72.69, 76.75, 81.30 and 88.42, respectively against *Alternaria solani* on potato dextrose agar medium using poison food technique.

Yadav *et al.* (2018) screened various fungicides and found that Hexaconazole @ 3000 ppm showed minimum radial growth (3.00 mm) of *Alternaria solani*, causing early blight of potato to evaluate the effect of different fungicide such as significantly at 3000 ppm concentration followed by Mancozeb (3.16 mm) as compare to control (15.03 mm).

Yadav *et al.* (2018) a study was carry out in the department of plant pathology, college of horticulture, Bharsar. Throughout *kharif* season 2016 to control *Alternaria solani* early blight of potato to evaluate the effect of different fungicide such as Hexaconazole and Mancozeb significantly lower the disease intensity (4.81, 10.51 and 18.26%) at 45, 60, 75 DAS and maximum tuber yield (446.95 g/plant) followed by Mancozeb (6.50, 13.70 and 22.00%) at 45, 60, 75 DAS and tuber yield (428.03 g/plant) as compare to control (20.00, 38.60 and 55.25%) at 45, 60, 75 DAS and (307.12 g/plant) *in vivo* condition.

Bansode *et al.* (2018) revealed that spray of Hexaconazole 5 EC @ 0.05% and then Mancozeb 75 WP @ 0.25% at 10 days interval was found significantly superior in controlling the early blight disease of potato with highest yield (22.40 t/ha) as compared to other treatments.

Kumar *et al.* (2018a) recorded that Propiconazole 25% EC (83.62%) gave maximum inhibition of the mycelial growth of *Alternaria solani* followed by Mancozeb 75% WP (56.85%) at 200 ppm concentration in early blight of potato in *vitro* condition.

Kumar *et al.* (2018a) a field experiment was conducted during *Rabi* season 2013 to control early blight of potato in Kufri Bahar variety to evaluate the Propiconazole 25% EC was most effective followed by Mancozeb 75% WP and also economical in reducing severity of the early blight and increasing yield over control.

Sharma *et al.* (2018) Evaluate the lowest per cent disease intensity (PDI) through the Difenconazole 25% EC @ 0.025% (20.59%) followed by Propiconazole 25% EC @ 0.025% (21.52%) and Mancozeb 75% WP @ 0.2% (24.72%) treatments as compare to untreated control on the early blight of potato.

Ahmad *et al.* (2019) revealed that Mancozeb 75 WP @ 2000 ppm gave lowest mycelial growth and highest inhibition percentage (3.65 mm and 85.53%) of *Alternaria solani* in potato.

Sreenivasunu *et al.* (2019) tested fungicides against *Alternaria solani* by using poison food technique and found that Propiconazole 25% EC at 0.1% gave 90.26% inhibition followed by Mancozeb 75% WP at 0.25% (87.57% inhibition). Similar trend of fungicide observed under field conditions, spray of Propiconazole 25% EC @ 0.1% gave highest yield (213.97 q/ha) followed by Mancozeb 75% WP at 0.25% concentration gave yield (178.20 q/ha). While, lowest yield (107.90 q/ha) in untreated control.

## Chapter 3

### MATERIAL METHOD

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Present investigation on “**Detection and management of early blight of potato caused by *Alternaria alternata***” was carried out at the field of Agricultural Research Station, Ummedganj and in laboratory of college of Agriculture, Agriculture University-Kota during the year 2019-20. The materials used and methods adopted during these studies are described here in this chapter:

#### 3.1 General

##### 3.1.1 Experimental site

The laboratory experiments were conducted at Laboratory of Plant Pathology, College of Agriculture, ARS, Kota and field experiments were conducted at the experimental site Agricultural Research Station, Ummedganj-Kota. Kota is situated at latitude 25.21° N, longitude of 75.86° E and altitude of 271 meters above MSL (mean sea level). The region falls under semi-arid eastern plain (Agro Climatic Zone- V) of Rajasthan.

##### 3.1.2 Climate and Weather Condition

The maximum mean daily temperature ranges from 24.5°C in the month of January and 42.6°C in May and minimum 10°C in January and 19.7°C in month of May.

##### 3.1.3 Soil

The soil of the University service area ranges from clay loam to clay. The major soils are Vertisols (71.7%), Alfisols (12.7%), and Inceptisols (4.8%). The average normal rainfall of the university jurisdiction varies 732-1005 mm and major source of irrigation are canal through rivers *viz.*, Chambal, Kalisindh and Parvanetc and tube wells.

#### 3.2 Raising of crop

The field was prepared before sowing by cross ploughing with tractor drawn disc harrow and planked. The field experiments were conducted on variety Kufri Bahar of potato in *Rabi* season 2019-20 except screening of genotypes. The crop was raised in plots of 3.0 × 2.4 m<sup>2</sup> keeping P × R distance of 60 × 20 cm<sup>2</sup>. The irrigation was applied as per the recommendations for the crop in this zone. One weeding and hoeing was done as per recommendation.

### **3.3 General laboratory procedure**

#### **3.3.1 Glassware cleaning**

For all the laboratory experimental studies, Corning and Borosil glassware's were used. The glassware's were kept in cleansing solution containing 60.0 g of potassium dichromate ( $K_2Cr_2O_7$ ) 60.0 ml of concentrated sulphuric acid ( $H_2SO_4$ ) in 1000 ml of water for 24 hours. They were washed with detergent powder followed by washing in running tap water and then rinsed in distilled water before use.

#### **3.3.2 Sterilization**

All glass wares, solid and liquid media were exposed to sterilization by autoclaving at  $1.1 \text{ kg/cm}^2$  ( $121.6^\circ\text{C}$ ) for 15 min. The seeds and plant tissues were surface sterilized in 1 per cent sodium hypochlorite solution for one minute followed by three changes in sterile water. All cultural studies were conducted under barren conditions in laminar flow. The tip of inoculated needle and forceps were disinfected over flame.

### **3.4 Preparation of media**

#### **3.4.1 Potato dextrose agar medium**

For all the laboratory experimental studies, standard potato dextrose agar (PDA) medium was used for culturing the *Alternaria alternata*.

The composition of PDA used is given below.

Peeled potato	:	200 g
Dextrose	:	20 g
Agar-agar	:	20 g
Distilled water	:	1000 ml

Two hundred grams of peeled potatoes were cut into pieces. These pieces were boiled in water and the extract was collected by filtering through muslin cloth. Each of 20 g of dextrose and agar-agar were dissolved in potato extract and the final volume was made up to 1000 ml by adding distilled water. A known quantity of such medium was dispensed into number of conical flasks and plugged with nasty cotton and finally wrapped with paper. The flasks containing provided medium were sterilized at a pressure of  $1.1 \text{ kg/cm}^2$  for 15 minutes.

### **3.5 Symptoms and isolation of the pathogen**

Early blight symptoms of potato were isolated from infested leaf sample of potato plants showing typical symptoms on Kufri Bahar variety. The diseased leaves were examined for associated fungus by teasing the diseased portion with the help of a teasing needle and was observed under microscope at the margins of the diseased spots (10x × 10x). For the isolation of fungus, small segments of diseased tissue along with some healthy leaf portion (5 × 5 mm<sup>2</sup>) was cut with a sterilized razor blade at the margins of the diseased spots on the leaves and surface sterilized in 0.1% mercuric chloride solution for 30 second. The leaf segments were rinsed thrice in distilled sterilized water to remove the last trace of mercuric chloride solution, blotted dry and placed on acidified potato dextrose agar medium (PDA) in sterilized Petri plates. Three pieces of sterilized specimen were placed in each Petri plate and incubated for 7 days at 25 ± 1°C. One set of PDA plates was seeded with bits without mercuric chloride treatment.

### **3.6 Purification of the pathogen**

The isolated pathogen was purified by single spore isolation and maintained on PDA slants. The mycelial growth was observed in Petri plates, advancing hyphal tips growing out of tissue segments were cut off with sterilized inoculation needle and transferred to potato dextrose agar slants for further growth in Petri plates and incubated at 25 ± 1°C for 24 h. The pure cultures thus obtained, was maintained by repeated sub-culturing at an interval of 30 days for further studies. The stock culture in PDA slants was stored at 4°C in refrigerator. The culture was purified and maintained for further studies.

### **3.7 Pathogenicity of pathogen**

To prove the ability of the isolated fungus to cause early blight disease in potato, were inoculated by spraying with conidial suspension of *Alternaria alternata*. The conidial suspension was prepared in sterilized distilled water by blending 7 days old fungal culture in grinder and mortar and filtered through cheese cloth. Conidia per ml were counted with the help of haemocytometer and were diluted with sterilized distilled water to have 1 × 10<sup>5</sup> conidia per ml of water. 15 days after transplanting potato plants were inoculated by spraying the conidial suspension with the help of an atomizer. The control was also maintained by spraying sterilized distilled water only. Inoculated and control plants were covered with plastic bags and incubated in a cage

house for 72 hours at 25°C, then uncovered and kept wet until symptoms appeared. One week after inoculation, small dark spots were observed, that rapidly developed into necrotic areas on the potato leaves. Re-isolation was done prepared from artificially inoculated plants and resulting culture was compared with the original one with admiration to colony characters, colony color, conidial morphology and septation.

### 3.8 Identification of pathogen

The morphological study was carried out. The observations were recorded from the 7 days old culture for the characters of colony, mycelium and conidia. The pathogen was identified on the basis of colony characters, viz., color, growth, pigmentation etc. The culture was sent for identification at species level at Indian Type Culture Collection, IARI for confirmation.

### 3.9 Media study

A modified method suggested by Koley and Mahapatra (2015) was followed to find favorable medium. Various leaf decoction media from plant species as well as synthetic and semi synthetic media as listed below were prepared and evaluated. Mycelial growth and dry mycelial weight was recorded. The broth media of below mentioned media prepared and dry mycelium weight was calculated (mg).

#### Experimental Details

Design :CRD Treatments :8 Replications :4

#### Treatment details

Sr. No	Name of media
1.	V-8 juice agar
2.	Czapek's agar
3.	Potato dextrose agar
4.	Oat meal agar
5.	Potato stem agar
6.	Potato leaf agar
7.	Potato carrot agar
8.	Pea agar

The composition and preparation of the above mentioned synthetic and non-synthetic/semi-synthetic media were obtained from Ainsworth and Bisby's "Dictionary of the Fungi" by Hawksworth *et al.* (1983).

The composition and preparation of the media is as follows:-

**1. V-8 juice agar**

V-8	:	200 ml
CaCO <sub>3</sub>	:	3.0 g
Agar- Agar	:	20 g
Distilled water	:	1000 ml

Suspend 44.3 g in 1000 ml of distilled water and boiling. Sterilize by autoclaving at 15 lbs pressure 121°C for 15 minutes. We made a solution of 4.43 g in 100 ml distilled water.

**2. Czapek's agar**

Sucrose	:	30.0 g
Sodium nitrate (NaNO <sub>2</sub> )	:	2.00 g
Potassium dihydrogen phosphate (KH <sub>2</sub> PO <sub>4</sub> )	:	1.00 g
Magnesium sulphate (MgSO <sub>2</sub> . 7H <sub>2</sub> O)	:	0.50 g
Ferric chloride (FeCl <sub>3</sub> . 6H <sub>2</sub> O)	:	0.01 g
Potassium chloride (KCl)	:	0.50 g
Agar - agar	:	20.00 g
Distilled water	:	1000 ml (volume to make up)

Agar-agar was melted in 500 ml distilled water. The other ingredients were dissolved in remaining 500 ml of distilled water. The two solutions were mixed thoroughly and the volume was made up to one litre. The medium was sterilized at 1.1 kg/cm<sup>2</sup> pressure for 15 minutes.

**3. Potato dextrose agar (PDA)**

Peeled potato	:	200.00 g
Dextrose	:	20.00 g
Agar-agar	:	20.00 g
Distilled water	:	Distilled water

200 gram of peeled potato was cut into small bits and boiled in distilled water and extract was collected by filtering through muslin cloth. Dextrose 20 g and Agar- agar powder 20 g each were dissolved in potato extract and the final volume was made up to 1000 ml with distilled water. Later, it was sterilized at 1.1 kg/cm<sup>2</sup> pressure for 15 minutes and preserved for future use. We made a solution of 3.9 g in 100 ml water.

#### 4. Oat meal agar

Oat meal powder	:	40.00 g
Agar - agar	:	20.00 g
Distilled water	:	1000 ml (volume to make up)

The oat meal powder was dissolved in 500 ml of distil water and agar- agar was melted in 500 ml of distilled water separately. Both the solutions were mixed thoroughly. Then the volume was made up to one litre and sterilized.

#### 5. Potato stem agar

Healthy potato plant stem	:	20.00 g
Agar-agar	:	2.00 g
Distilled water	:	100 ml

Potato plant stem were boiled in 50 ml water for 30 minutes. Extract was collected by filtering through muslin cloth. The agar-agar was melted in 50 ml water, both the solutions were mixed and the volume was made up to 100 ml and was sterilized.

#### 6. Potato leaf Agar

Healthy potato plant leaves	:	20.0 g
Agar-agar	:	2.00 g
Distilled water	:	100 ml

Potato plant leaves were boiled in 50 ml water for 30 minutes. Extract was collected by filtering through muslin cloth. The agar-agar was melted in 50 ml water, both the solutions were mixed and the volume was made up to 100 ml and was sterilized.

#### 7. Potato carrot agar

Carrot	:	200 g
Potato	:	250.00 g
Agar-agar	:	15.00 g
Distilled water	:	1000 ml

Carrots were boiled for 10-15 minutes in 400 ml distilled water; the extract was squeezed and filtered through muslin cloth. The extract was collected in a beaker, remaining ingredients were added to same and volume was made to one litre before sterilization.

#### 8. Pea agar

Pea	:	20.00 g
Agar-agar	:	2.00 g
Distilled water	:	100 ml

Pea (Green) was boiled in 100 ml water for 30 minutes. Extract was collected by filtering through muslin cloth. The agar-agar was melted in 100 ml water and the volume was made up to 100 ml and was sterilized.

The sporulation of the spore suspension was estimated as per (Krishna *et al.*, 2018)

<b>Number of spore per microscopic field</b>	<b>Designation</b>
0	- (nil)
1-10	+ (poor)
11-20	++ (moderate)
21-30	+++ (good)
31-40	++++ (excellent)

### **3.10 Efficacy of bio-agent against the pathogen *in-vitro* condition**

Various known bio-agent *Trichoderma* species viz., *Trichoderma virens*, *Trichoderma azospirillum*, *Trichoderma harzianum*, *Trichoderma viride* were collected from Department of Plant Pathology and screened for their effectiveness against *Alternaria alternata* by using CRD for their effectiveness using dual culture technique.

#### **Experimental Details**

Design : CRD  
Treatment : 5  
Replications : 4

#### **Treatment details of bio-control agents**

<b>S. No.</b>	<b>Name</b>
1	<i>Trichoderma virens</i>
2	<i>Trichoderma azospirillum</i>
3	<i>Trichoderma harzianum</i>
4	<i>Trichoderma viride</i>
5	Control

#### **Dual culture technique**

Bio-control agents were evaluated amount for their efficacy against *Alternaria alternata* using dual culture technique. Twenty ml of potato dextrose agar was poured into 90 mm diameter Petri dishes and permit to solidify. Mycelial disc of (6 mm) from seven days old actively growing culture of the bio-agents and the test pathogen were cut separately with the help of sterilized cork borer and placed on solidified PDA

approximately, 4 cm away from each other. Each treatment was replicated five times and incubated at  $25 \pm 1^\circ\text{C}$ . The activity of antagonistic organisms were recorded by measuring the colony diameter of *A. alternata* in each treatment and compared with control. The per cent inhibition of growth of the pathogen was calculated by using the formula suggested by Vincent (1947).

$$I = \frac{C-T}{C} \times 100$$

Where

I= Per cent inhibition of mycelium

C= Growth of mycelium in control

T= Growth of mycelium in treatment

### 3.11 Screening the genotypes against the disease

Potato genotypes were screened against the early blight disease under natural conditions. From each genotype, 10 plants were selected randomly and kept unsprayed throughout the season and tagged for the assessment of the disease. Twenty eight genotypes as listed below were evaluated for their reaction to early blight disease of potato. Disease rating was done on a 0-5 arbitrary scale (Horsefall and Barette, 1945).

#### Description of the symptoms and disease reaction

Category	Description of the symptoms	Reaction
0.	Leaves free from infection	HR
1.	Small irregular spots covering <5% leaf area	R
2.	Small irregular brown spots with concentric rings covering 5.1-10% leaf area	MR
3.	Lesions enlarging, irregular brown with concentric rings covering 10.1-25% leaf area	MS
4.	Lesions coalesce to form irregular and appears as a typical blight symptoms covering 25.1-50% leaf area	S
5.	Lesions coalesce to form irregular and appears as a typical blight symptoms covering >50% leaf area	HS

HR= highly resistant, R= resistant, MS= moderately susceptible, S= susceptible, HS= highly susceptible.

**Name of genotypes/varieties**

<b>Sr. No.</b>	<b>Name of genotypes/varieties</b>
1.	NJ-1501
2.	MP-97
3.	MP/94-322
4.	NJ-44
5.	NJ-85
6.	NJ-34
7.	MP-97-1606
8.	NJ-1530
9.	PS/RI-135
10.	CP-3021
11.	J-92-164
12.	JI-93-77
13.	MS/94-118
14.	JN-1177
15.	MS/92-2105
16.	J/93-4
17.	NJ-1
18.	MS/85-1663
19.	Kufri Sindhuri
20.	Kufri Bahar
21.	Kufri Pukhraj
22.	Kufri Khayati
23.	Kufri Pushkar
24.	3153
25.	Atlantic
26.	Chip- 3
27.	Jx-161
28.	Ms/78-62

**The disease incidence was calculated by using the following formula**

$$\text{Per cent disease incidence} = \frac{\text{No. of diseased leaves}}{\text{Total No. of leaves examined}} \times 100$$

### **3.12 In vitro evaluation of fungicides**

Various fungicides as listed below at different concentrations were evaluated for their effectiveness against pathogen by poisoned food technique as suggested by Nene and Thapliyal (1979).

The efficacies of systemic, non-systemic and products were assessed by poison food technique. The pathogen *Alternaria alternata* was cultured on PDA medium in Petri plates for ten days before setting the experiment. The details are mentioned below:-

#### **3.12.1 Poisoned food technique**

Fungicide suspension was prepared in PDA by adding required quantity of fungicide to obtain the desired concentration on the basis of active ingredient and whole product present in the chemical. Twenty ml of poisoned medium was poured in each of the sterilized Petri plates. Mycelial disc of 0.5 cm was taken from the outer side of ten days old culture and was placed in the center and incubated at  $25 \pm 1^\circ\text{C}$  till growth of the fungus touched the periphery in control plate. Suitable checks also maintained without addition of any fungicide. Four replications were maintained for each treatment. The diameter of the colony was measured bidirectional and average was worked out. The per cent inhibition of growth was calculated by using the formula given by Vincent (1947).

$$I = \frac{C-T}{C} \times 100$$

Where

I= Per cent inhibition of mycelium

C= Growth of mycelium in control

T= Growth of mycelium in treatment

#### **Experimental Details**

Design : CRD Replications : 4 Treatments : 8

### Treatment of fungicides and concentration

Sr. No.	Name of fungicides	Concentration (ppm)
1.	Difenoconazole 25% EC	10, 20, 50
2.	Propiconazole 25% EC	10, 20, 50
3.	Hexaconazole 5% EC	10, 20, 50
4.	Azoxystrobin 23% SC	20, 50, 100
5.	Picoxystrobin 22.52 SC	20, 50, 100
6.	Pyraclostrobin 20% WG	20, 50, 100
7.	Mancozeb 75% WP	500, 1000, 1500
8.	Control	

#### 4.13 *In vivo* evaluation of fungicides

A field experiment on potato crop was conducted at Agricultural Research Station, Kota district during *Rabi* 2019-20 to know the efficacy of all the above mentioned fungicides. The experiment was laid out in Randomized Block Design (RBD) with three replications. Potato tubers were sown on 7th November 2019 and harvested on 29 February 2020. Two sprays of the fungicides were given at 15 days interval. The first spray was given immediately after the first appearance of early blight symptoms i.e. 45 days after planting. Second spray was given at 60 days after planting. Five plants in each subplot were considered for disease and data were converted into per cent disease intensity (PDI) and per cent disease incidence as explained earlier. Finally yield data was taken after harvest.

#### Treatment details of fungicides with dose (g) a.i./ha

S. No.	Name of fungicides	Dosage (ml/g)/ha		
		<i>a.i.</i>	Formulation(ml)	Water (L)
1.	Difenoconazole 25% EC	50 ml	250	500
2.	Propiconazole 25% EC	125 ml	500	500
3.	Hexaconazole 5% EC	25 ml	500	500
4.	Azoxystrobin 23% SC	125 ml	500	500
5.	Picoxystrobin 22.52 SC	100 ml	400	500
6.	Pyraclostrobin 20% WG	100 g	500	500
7.	Mancozeb 75% WP	2250 g	1500	500
8.	Control (only water spray)	-	-	-

**The disease intensity was calculated by using the following formula**

$$\text{Per cent Disease Intensity} = \frac{\text{Sum of Individual rating}}{\text{Number of Plant Observed} \times \text{Maximum Disease rating}} \times 100$$

**The per cent disease control was calculated by using the following formula**

$$\text{Per cent disease control} = \frac{\text{Disease in control} - \text{Disease in treatment}}{\text{Disease in control}} \times 100$$

**The disease incidence was calculated by using the following formula**

$$\text{Per cent disease Incidence} = \frac{\text{No. of diseased leaf}}{\text{Total number of leaves examined}} \times 100$$

**The yield increase over control was calculated by using the following formula**

$$\text{Yield increase over control} = \frac{T-C}{C} \times 100$$

Where,

T = Yield of respective treatment (kg/ha)

C = Yield of control (kg/ha)

### **3.14 Statistical analysis**

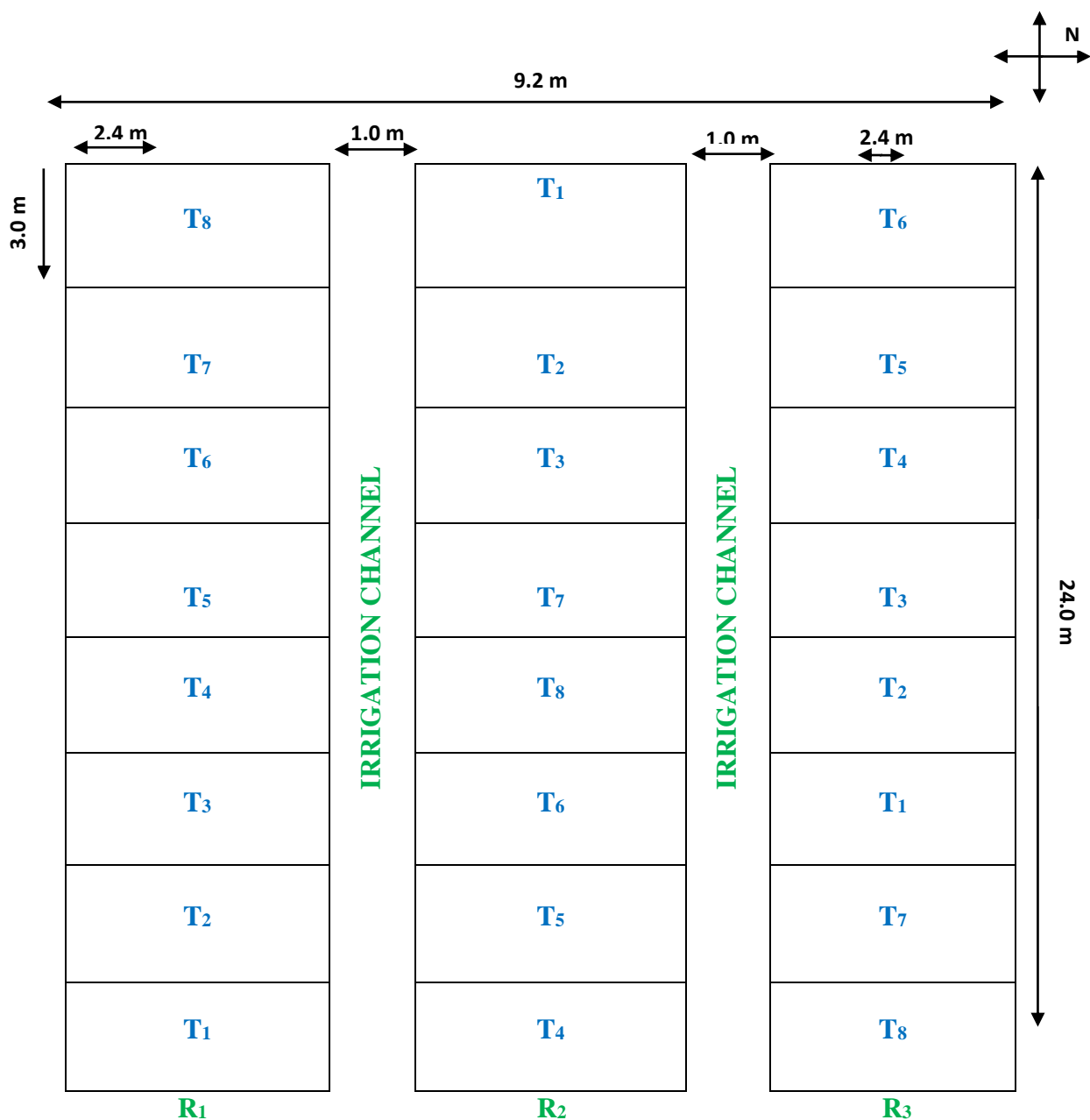
Randomized Block Design (RBD) (for cultivation of crop early blight incidence and yield) and Completely Randomized Design (CRD) (for *in vitro* experiments) were employed to analyse the data. The critical differences were worked out at 5% probability level. The experiments were conducted in the PG laboratory, Department of Plant Pathology, College of Agriculture, Ummedganj-Kota. Agriculture Research Station Ummedganj-Kota.

### **3.15 Cost benefit ratio**

The cost benefit ratio was calculated to see the total benefit gained by using the fungicides i.e. score one application, score two application.

Cost benefit ratio was computed by using the formula

$$B: C = \frac{\text{Gross Income}}{\text{Net income}}$$



### Treatment details

T <sub>1</sub> -	Difenoconazole 25% EC
T <sub>2</sub> -	Propiconazole 25% EC
T <sub>3</sub> -	Hexaconazole 5% EC
T <sub>4</sub> -	Azoxystrobin 23% SC
T <sub>5</sub> -	Picoxystrobin 22.52 SC
T <sub>6</sub> -	Pyraclostrobin 20% WG
T <sub>7</sub> -	Mancozeb 75% WP
T <sub>8</sub> -	Control

### Other experimental details

- i. Design - RBD
- ii. Treatment - 8
- iii. Replication – 3
- iv. Plot size – 3 m x 2.4 m
- v. Spacing- 60 cm x 20 cm
- vi. Spacing between replication - 1.0 m
- vii. Date of sowing- 7.11.2019
- viii. Date of harvesting- 29.02.2020



## Chapter 4

### EXPERIMENTAL RESULTS

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The present research entitled “**Detection and management of early blight of potato caused by *Alternaria alternata***” was conducted at the field of Agriculture Research Station, Ummadganj and in laboratory of college of Agriculture, Agriculture University-Kota, Rajasthan. The data recorded on different aspects and the results interpreted are presented in this chapter as under:

#### **4.1 To study the symptoms, isolation, purification, pathogenicity and identification of pathogen**

##### **4.1.1 Symptomatology**

Symptomatology study aids in providing information about symptoms produced by early blight disease and helps for diagnosis of disease in field conditions. Symptoms of early blight disease vary with potato variety, environmental conditions and stage of plant growth at the time of infection. The disease plants under field were observed weekly starting from germination to harvesting of crops. During the course of study disease symptoms become visible on all plant parts *viz.*, leaf, stem, petioles and tubers. The first symptom of the disease was seen on older leaves as small dark brown, frequently round necrotic spots of 1-2 mm diameters later, the spots enlarged with characteristics in center to produce a target board effect and the colour of spots altered from brown to dark brown. The contiguous spots eventually coalesce to form large irregular spots primary to drying and defoliation. When plants were 60-90 days mature, symptoms also appeared on stem and petioles as brown extended cankerous target broad spots. These enlarged and covered whole stem and petioles leading to withering of plants. The diseased plants produced less number of tubers and small sized tuber as compared to healthy plant. (Plate 4.1)

##### **4.1.2 Isolation and purification**

The associated pathogen was isolated by single spore isolation technique from infected potato leaves and pure culture of the fungus was obtained on potato dextrose agar medium. The pure culture of the fungus was obtained after eight days of inoculation which showed brownish colour. Purity and virulence of isolated fungus was regularly maintained and monitored by sub-culturing after 30 days and the culture

was maintained at 5°C. For further studies, the original culture was revived once in 30 days (Plate 4.2).

#### **4.1.3 Pathogenicity of the fungus**

Pure culture was obtained by single spore isolation method and such culture was used for pathogenicity test. Inoculation with spore suspension and homogenized mycelial piece of *A. alternata* on foliage of 55 days old potato plants. The result revealed that the primary symptoms of disease occur on light brown spots seven days after inoculation as small chlorotic, disintegrate and light brown spots. Re-isolated and purified culture from these artificially infected leaves was similar to that of original culture in provided environmental conditions during study. The plants which were not inoculated with the fungal spore suspension did not show any symptoms of the disease (Plate 4.3 and 4.4).

#### **4.1.4 Identification of the pathogen**

An associated fungus was identified on the basis of its morphological and colony characters. The mycelium was greyish brown to black producing conidiophores moreover singly or in small groups which were straight or flexuous brown to olivaceous brown. The conidia were solitary straight or muriform or ellipsoidal narrowing to beak, pale or olivaceous brown, length 36.15-26.85 µm and 18.65-9.92 µm thick at the broadest piece with 3-4 transverse and 1-3 longitudinal septa. The beaks were flexuous, pale and occasionally branched. The identification of *Alternaria alternata* was further confirmed from Indian Type Culture Collection, Division of Plant Pathology, IARI, New Delhi-110012 with code number 295012.

### **4.2 To study the efficacy of bio-agents and media study against the pathogen *in vitro* conditions**

#### **4.2.1 Efficacy of media study**

To find out the best medium for the mycelial growth and dry mycelial weight of *Alternaria alternata*, eight different solid and broth media were selected. The results presented in Table 4.1, Fig. - 4.1, and Plate 4.5 revealed that pathogen grew well on all the media tried. Perusal of data revealed that potato leaf agar medium was



Small irregular to circular dark brown spots



Concentric ring giving “target spot” or “bull eye” appearance



Narrow yellow halo around each spot



Severely infected leaf

Plate – 4.1: Symptoms showing infected plants leaves



Plate – 4.2: Pure culture and spores of *A. alternata*





Plate – 4.3: Spraying of conidial suspension on potato plants for pathogenicity

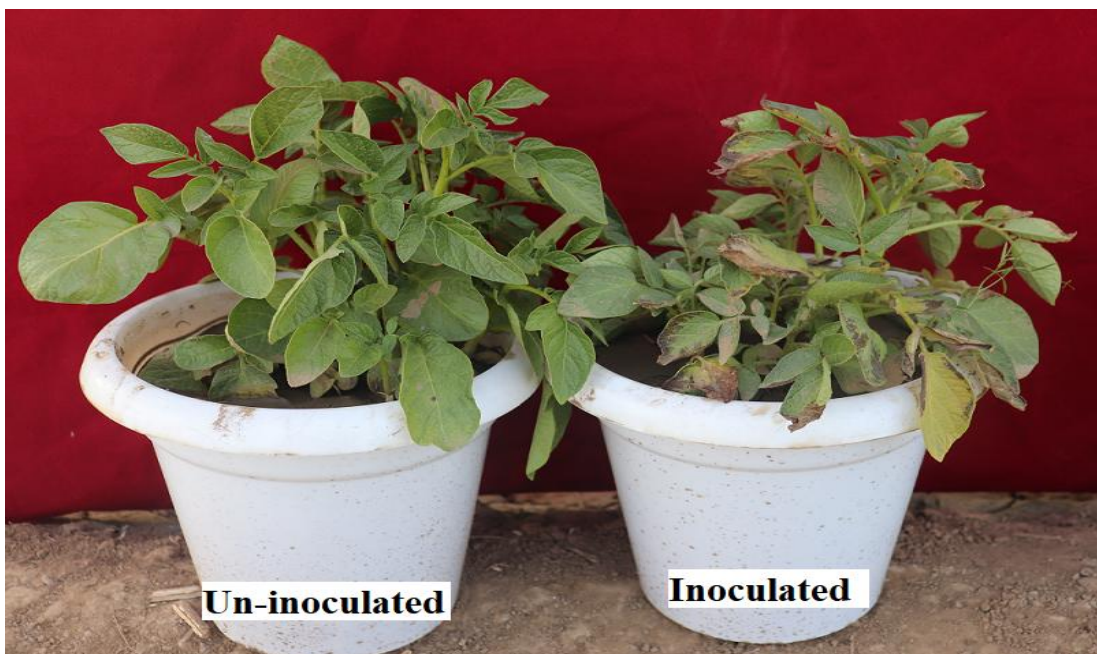


Plate – 4.4: Proving the pathogenicity of early blight of potato in pots



significantly superior in supporting maximum mycelial growth at 5 days (70.50 mm) and 7 days (80.25 mm) of after inoculation the pathogen. This was followed by potato dextrose agar medium at 5 days with mycelial growth (64.75 mm) and 7 days (78.25 mm) over all other media used.

The next best media for mycelial growth in order of merit were potato stem agar medium, V-8 juice agar medium and pea agar medium with growth diameter at 5 days and 7 days (59.00, 57.50 and 58.00 mm), (67.25, 64.50, and 63.25 mm) respectively, three being statistically at par with each other. However, oat meal agar medium and potato carrot agar medium producing growth at 5 days and 7 days (50.00 and 46.50 mm), (58.25 and 53.75 mm), respectively and these both media were statistically at par with the each other. While, the czapek's agar medium supported least growth diameter of the fungus at 5 days and 7 days (37.50 and 42.75 mm).

Maximum dry mycelium weight i.e. 584.75 mg was recorded on potato leaf broth medium which was significantly superior as compare to other media used in study. Next best dry mycelium weight was recorded on potato dextrose broth 570.25 mg. The next in order of merit were potato stem broth 315.50 mg, V-8 juice broth 285.00 mg, pea broth medium 268.50 mg, oat meal broth 244.25 mg, potato carrot broth 212.25 mg while, and minimum in czapek's broth 176.50 mg (Plate – 4.6 and Fig.– 4.2).

Among the media tested, the potato leaf agar was significantly superior and most suitable medium for density of mycelial as well as dry mycelial weight followed by PDA.

The test culture media exhibited a wide range of sporulation. However, potato leaf agar exhibited excellent (++++) sporulation. It was good (+++) on potato dextrose agar and potato stem agar; moderate (++) on V-8 juice agar and potato carrot agar; whereas, it was poor (+) on Czapek's agar, Oat meal agar and pea agar medium. Whereas, the potato stem agar, V-8 juice agar, pea agar, oat meal agar, potato agar and czapek's agar medium were least effective in mycelial density.

**Table 4.1 *In-vitro* evaluation of different media for mycelial growth of *A. alternata***

<b>T. No.</b>	<b>Treatment name</b>	<b>Mycelial growth at 5 days (mm)</b>	<b>Mycelial growth at 7 days (mm)</b>	<b>Dry mycelium weight in broth media (mg)</b>	<b>Sporulation</b>
1	V-8 juice agar	57.50 (49.31)*	64.50 (53.44)	285.00	++
2	Czapek's agar	37.50 (37.31)	42.75 (40.83)	176.50	+
3	Potato dextrose agar	64.75 (53.60)	78.25 (62.21)	570.25	+++
4	Oat meal agar	50.00 (45.00)	58.25 (49.76)	244.25	+
5	Potato stem agar	59.00 (50.19)	67.25 (55.10)	315.50	+++
6	Potato leaf agar	70.50 (57.12)	80.25 (63.63)	584.75	++++
7	Potato carrot agar	46.50 (42.99)	53.75 (47.16)	212.25	++
8	Pea agar	58.00 (49.62)	63.25 (52.69)	269.50	+
S. Em. ±		(0.86)	(0.89)	2.73	
CD at 5%		(2.50)	(2.60)	7.96	

**+ = Poor, ++ = Moderate, +++ = Good, ++++ = Excellent**

\*Figures in parentheses are arcsine transformed values

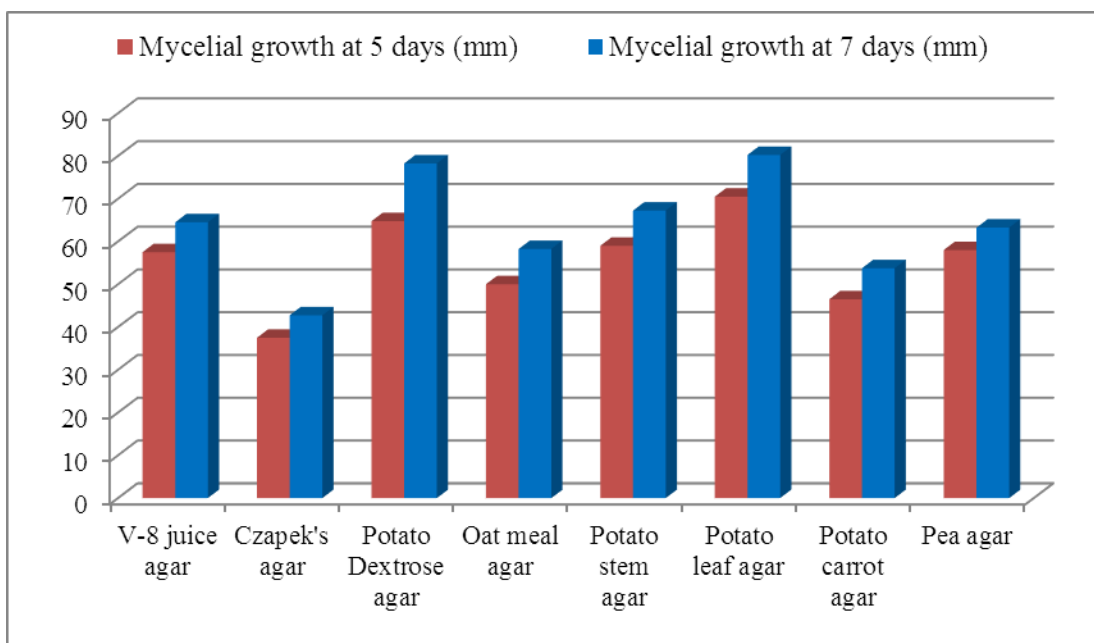


Fig. – 4.1: Evaluation of different solid media for mycelial growth of *A. alternata in vitro*

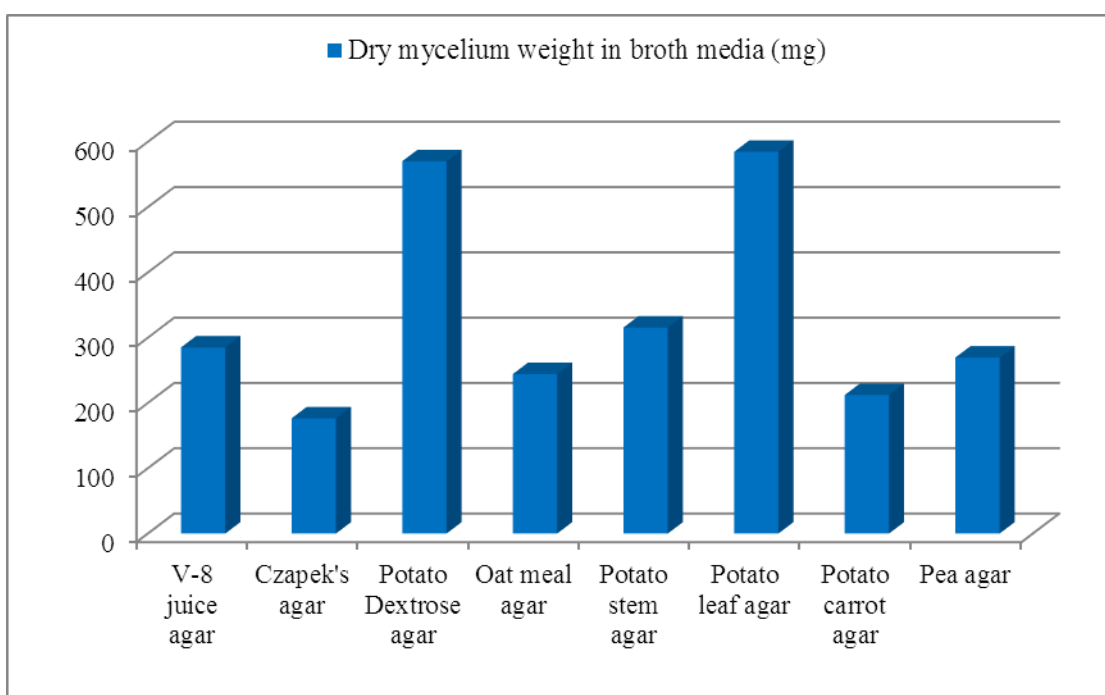
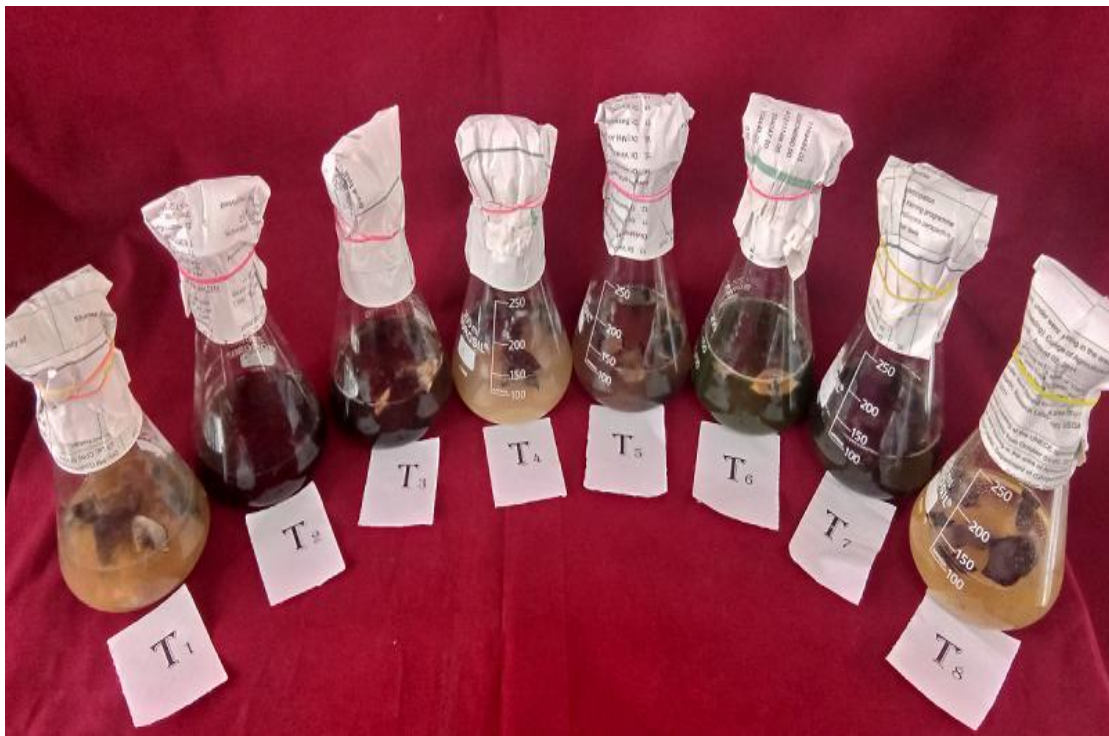


Fig. – 4.2: Evaluation of different broth media for dry mycelium weight of *A. alternata in vitro*





Plate – 4.5: Efficacy of different culture media against *A. Alternata*



**T<sub>1</sub>- V-8 Juice agar, T<sub>2</sub>- Czapek's agar, T<sub>3</sub>- Potato dextrose agar, T<sub>4</sub>- Oat meal agar, T<sub>5</sub>- Potato stem agar, T<sub>6</sub>- Potato leaf agar, T<sub>7</sub>- Potato carrot agar, T<sub>8</sub>- Pea agar**

Plate – 4.6: Dry mycelium weight of *A. alternata* on broth media



#### 4.2.2 Efficacy of bio-agents

Four bioagents of *Trichoderma* species viz., *Trichoderma virens*, *Trichoderma azospirillum*, *Trichoderma harzianum* and *Trichoderma viride* were evaluated for their antagonistic activity against *A. alternata* by dual culture technique. Based on the observations of radial growth of antagonist and fungus, per cent inhibition was calculated.

The results pertaining to antagonistic activity are presented in Table 4.2 revealed that all the four *Trichoderma* species significantly reduced the mycelial growth of the pathogen over the control. Significantly maximum inhibition of mycelial growth of the pathogen after 48 hours of incubation was obtained with *T. viride* (57.08%) followed by *T. azospirillum* (50.00%), *T. harzianum* (43.26%) and *T. virens* (37.08%) (Plate- 4.7 and Fig. - 4.3).

**Table 4.2 Effect of bio-agents against *A. alternata* in vitro by dual culture**

T. No	Treatment name	Mycelial growth At 48 hrs. (mm)	Per cent growth Inhibition
1	<i>Trichoderma virens</i>	28.00 (31.94)*	37.08
2	<i>Trichoderma azospirillum</i>	22.25 (28.14)	50.00
3	<i>Trichoderma harzianum</i>	25.25 (30.16)	43.26
4	<i>Trichoderma viride</i>	19.02 (26.02)	57.08
5	Control	44.50 (41.84)	--
S. Em. $\pm$		(0.47)	
CD at 5%		(1.41)	

\*Figures in parentheses are arcsine transformed values



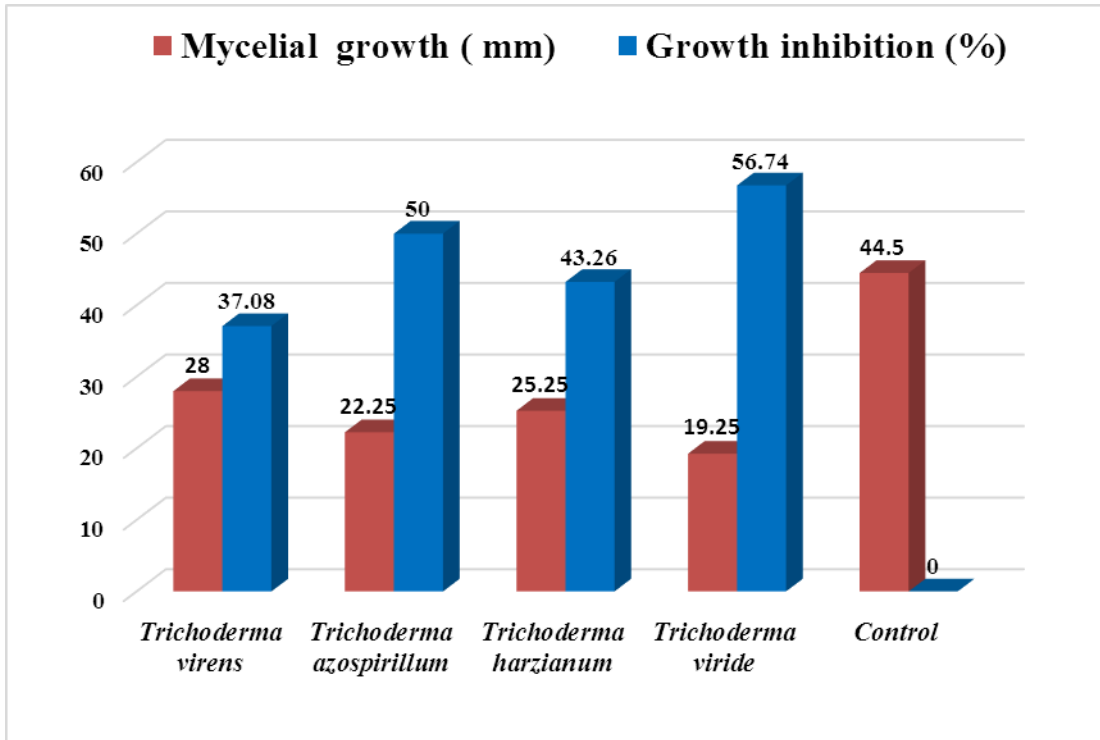


Fig. – 4.3: Effect of bio-agents against *A. alternata* in vitro condition



Plate – 4.7: In vitro evaluation of bio-agents against *A. alternata*



### 4.3 Screening the genotypes against the disease

Twenty-eight genotypes/varieties of potato were screened for resistance to early blight under natural field condition during *Rabi* 2019-20. The per cent germination as well as per cent disease incidence was calculated. The observation of early blight incidence was recorded at 60 and 75 days after sowing. Result in Table 4.3 and 4.4 revealed that different varieties were showed good germination percentage, which showed uniform growth of the plants (Plate - 4.8 and Fig. - 4.4).

Out of twenty eight genotypes/varieties, six genotypes *viz.*, CP-3021, 3153, Atlantic, Chip-3, Jx-161 and MS/78-62 showed resistant reaction, while thirteen genotypes *i.e.*, NJ-1501, MP-97, NJ-44, NJ-34, MP-97-1606, PS/RI-135, JI-93-77, MS/94-118, JN-1177, Kufri Sindhuri, Kufri Pukhraj, Kufri Khayati and Kufri Pushkar were moderately resistant. However, six genotypes *viz.*, NJ-85, NJ-1530, J-92-164, MS/92-2105, J/93-4 and Kufri Bahar were susceptible while, three genotypes *i.e.*, MP/94-322, NJ-1 and MS/85-1663 were highly susceptible.

**Table 4.3 Result summary of genotypes**

Categories	No.	Genotypes/varieties
Resistant	6	CP-3021, 3153, Atlantic, Chip-3, Jx-161 and MS/78-62
Moderate resistant	13	NJ-1501, MP-97, NJ-44, NJ-34, MP-97-1606, PS/RI-135, JI-93-77, MS/94-118, JN-1177, Kufri Sindhuri, Kufri Pukhraj, Kufri Khayati and Kufri Pushkar
Susceptible	6	NJ-85, NJ-1530, J-92-164, MS/92-2105, J/93-4 and Kufri Bahar
Highly susceptible	3	MP/94-322, NJ-1 and MS/85-1663

**Table 4.4 Screening of different genotypes/varieties against early blight of potato**

S. No.	Name of genotypes /varieties	Germination (%)	Disease incidence (%)	
			At 60 DAS	At 75 DAS
1	NJ-1501	75	8	17
2	MP-97	66	6	15
3	MP/94-322	87	38	52
4	NJ-44	80	6	13
5	NJ-85	86	25	38
6	NJ-34	80	10	18
7	MP-97-1606	83	7	16
8	NJ-1530	70	24	37
9	PS/RI-135	92	5	14
10	CP-3021	85	2	4
11	J-92-164	82	26	39
12	J1-93-77	73	6	17
13	MS/94-118	68	5	14
14	JN-1177	65	7	16
15	MS/92-2105	90	25	36
16	J/93-4	80	27	39
17	NJ-1	76	40	54
18	MS/85-1663	80	39	55
19	Kufri Sindhuri	86	9	18
20	Kufri Bahar	88	29	38
21	Kufri Pukhraj	80	8	15
22	Kufri Khayati	70	5	13
23	Kufri Pushkar	73	5	12
24	3153	82	1	3
25	Atlantic	87	3	5
26	Chip-3	85	1	2
27	Jx-161	79	2	3
28	MS/78-62	90	2	4

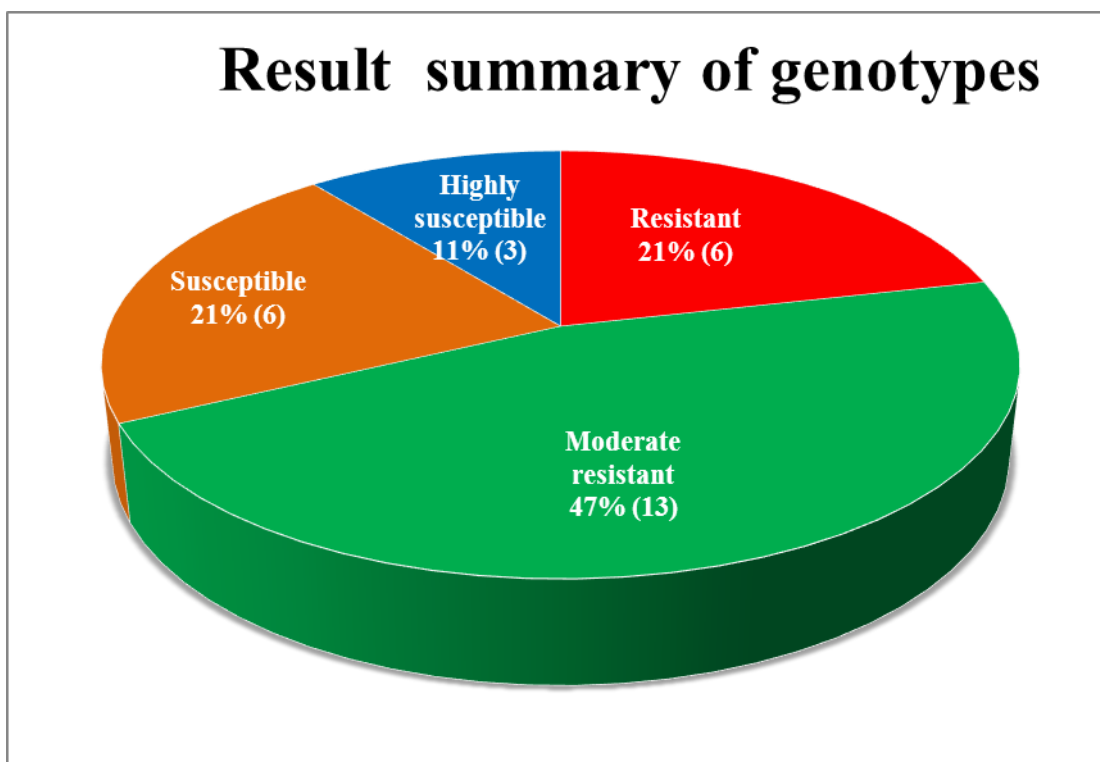


Fig. – 4.4: Result summary of genotypes under field condition



Plate – 4.8: Field view of potato genotypes



#### **4.4 To study the bio-efficacy of fungicide against the pathogen *in vitro* and *in vivo* conditions**

##### **4.4.1 Bio-efficacy of fungicides against the pathogen *in vitro***

The efficacy of various systemic as well as non-systemic fungicides *viz.*, Difenconazole, Propiconazole, Hexaconazole, Azoxystrobin, Picoxystrobin, Pyraclostrobin and Mancozeb were evaluated for their efficacy against *A. alternata* *in vitro* by poisoned food technique.

The results presented in (Table 4.5 and Plate 4.9) indicated that different fungicides have different efficacy against the growth of *Alternaria alternata* and all the fungicides significantly reduced the growth of the *A. alternata* as compare to control. Propiconazole 25% EC and Hexaconazole 5% EC both at all the three concentrations as well as Mancozeb 75% WP at 1000 and 1500 ppm and Difenconazole 25% EC at 50 ppm were found significantly statistically at par with 100 percent inhibition as compared to rest of the fungicides.

Next best treatments in order of merit were Difenconazole 25% EC at 10 and 20 ppm, Mancozeb 75% WP at 500 ppm and Azoxystrobin 23% SC at all the three concentrations significantly recorded minimum mycelium growth and maximum inhibition compared to control. However, Picoxystrobin 22.52% SC and Pyraclostrobin 20% WG both at all the three concentrations gave significantly maximum mycelium growth and minimum percent growth inhibition.

Present study revealed that Triazole group fungicide *viz.*, Propiconazole 25% EC, Hexaconazole 5% EC and Defenconazole 25% EC were significantly superior and proved the most effective in inhibiting mycelial growth @ 10, 20 and 50 ppm as compared to strobilin group *viz.*, Azoxystrobin 23% SC, Picoxytstrobin 22.52% SC and Pyraclostrobin 20% WG at 20, 50, 100 ppm against the *Alternaria alternata* *in vitro* condition by poison food technique. While contact fungicide Mancozeb 75% WP showed maximum inhibition at higher dose 1000 and 1500 ppm. (Fig. - 4.5a, 4.5b and 4.5c).

**Table 4.5 Effect of different fungicides and their concentrations on growth of *A. alternata* in vitro at 25 ± 1°C**

T. No.	Treatment	Concentration (ppm)	Mycelial growth (mm)	Percent inhibition
1	Difenconazole 25% EC	10	13.00 (21.11)*	85.56
		20	7.75 (16.14)	97.73
		50	0.00 (0.00)	100
2	Propiconazole 25% EC	10	0.00 (0.00)	100
		20	0.00 (0.00)	100
		50	0.00 (0.00)	100
3	Hexaconazole 5% EC	10	0.00 (0.00)	100
		20	0.00 (0.00)	100
		50	0.00 (0.00)	100
4	Azoxystrobin 23% SC	20	23.75 (29.16)	73.61
		50	14.50 (22.37)	83.89
		100	13.00 (21.12)	85.56
5	Picoxystrobin 22.52% SC	20	36.00 (36.87)	60
		50	32.25 (34.60)	64.17
		100	11.50 (19.80)	87.22
6	Pyraclostrobin 20% WG	20	41.50 (40.10)	53.89
		50	34.75 (36.12)	61.39
		100	12.50 (20.69)	86.11
7	Mancozeb 75% WP	500	11.50 (19.80)	87.22
		1000	0.00 (0.00)	100
		1500	0.00 (0.00)	100
8	Control		90.00 (71.56)	--
S. Em. ±			(0.39)	--
CD at 5 %			(1.11)	--

\*Figures in parentheses are arcsine transformed values

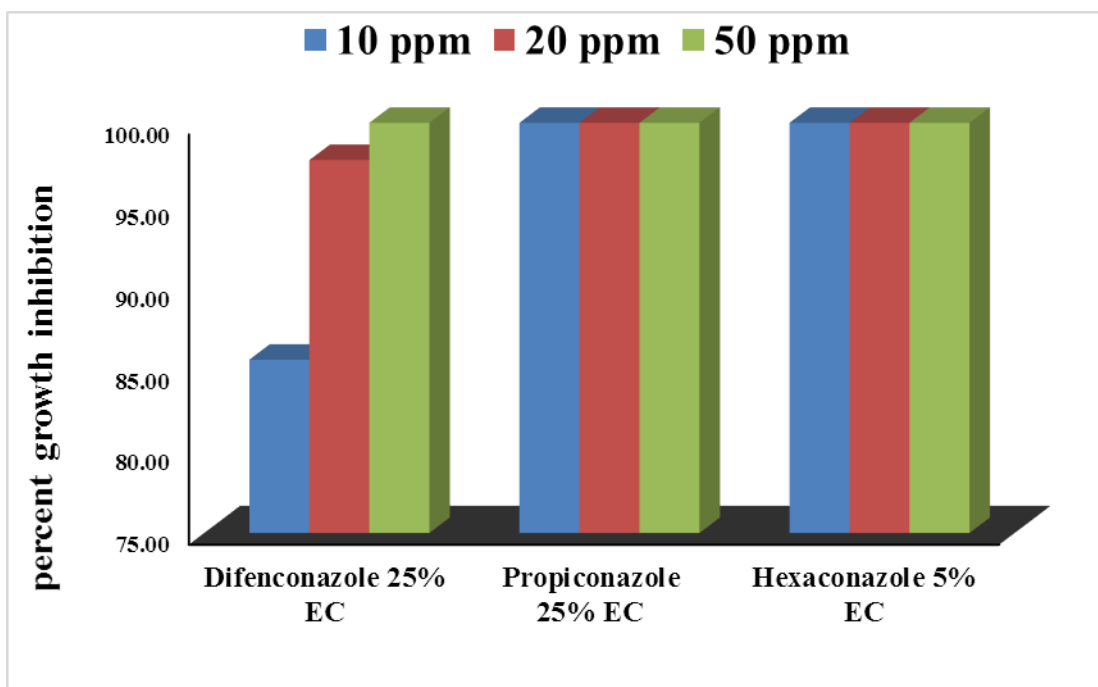


Fig. – 4.5a: *In vitro* bio-efficacy of trizole fungicides on percent growth inhibition of *A. alternata* by poisoned food technique at 10, 20 and 50 ppm concentrations.

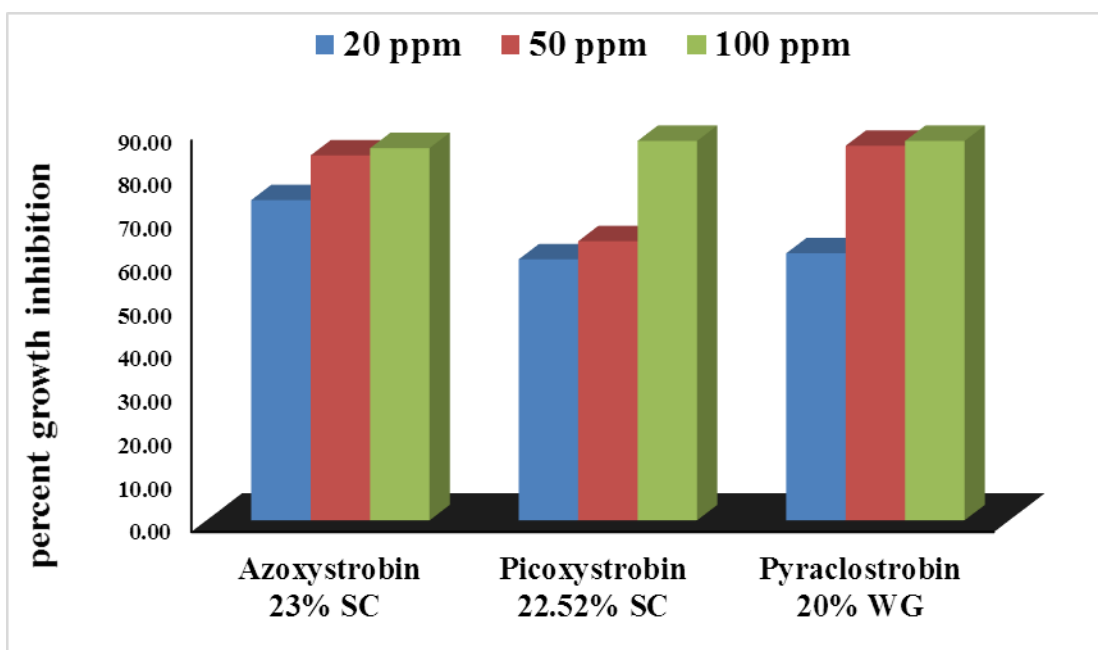


Fig. – 4.5b: *In vitro* bio-efficacy of strobilurin fungicides on percent growth inhibition of *A. alternata* by poisoned food technique at 20, 50 and 100 ppm concentrations.



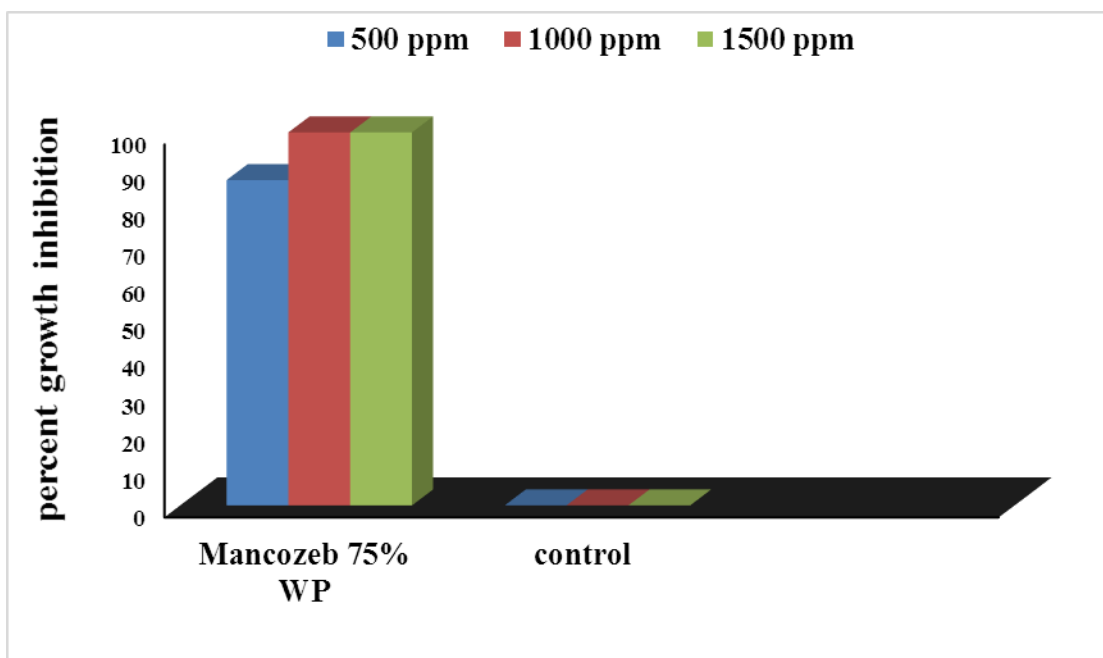


Fig. – 4.5c: *In vitro* bio-efficacy of contact fungicides on percent growth inhibition of *A. alternata* by poisoned food technique at 500, 1000 and 1500 ppm concentrations.

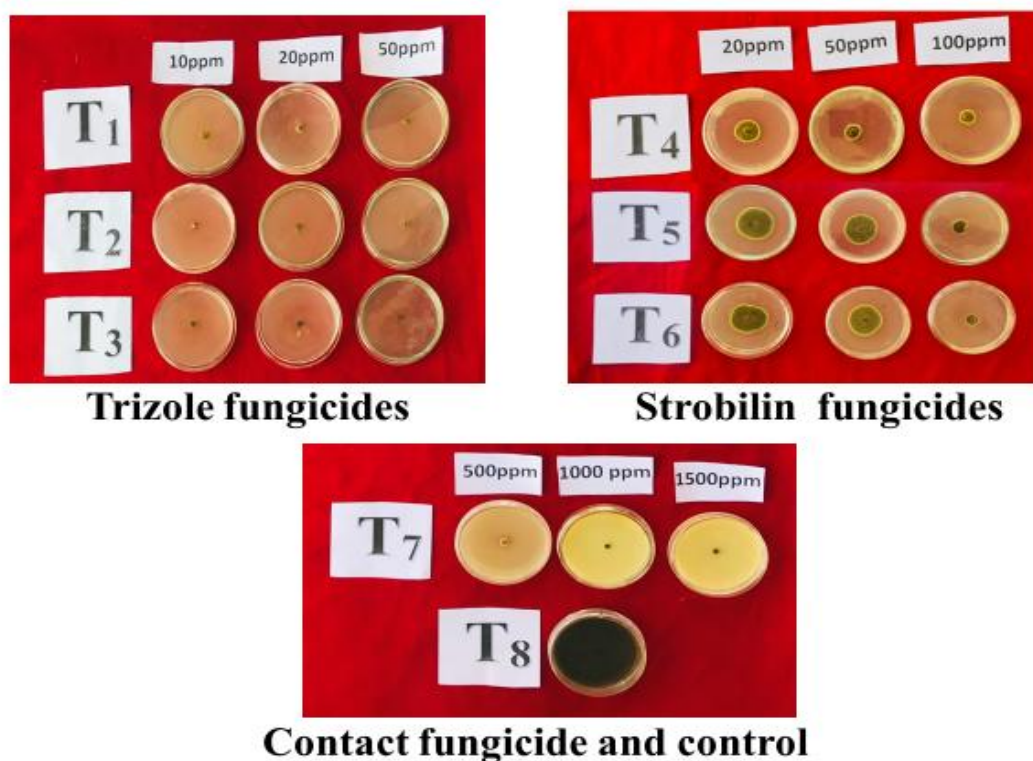


Plate – 4.9: Bio- efficacy of different fungicides against *A. alternata* *in vitro* condition



## 4.4.2 The bio-efficacy of fungicides

### 4.4.2.1 Per cent disease intensity

Fungicides found promising *in vitro* were further evaluated *in vivo* against early blight of potato. The germination was found non-significant in experiment. The first spray of the fungicide was given at initiation of early blight disease. All the tested systemic as well as non-systemic fungicides were found effective as foliar sprays against early blight of potato under field conditions and reduced the present disease intensity and gave significant reduction in disease over control. Significantly, minimum disease intensity was recorded in Dinfenconazole 5% EC @ 250 ml/ha after first (PDI 4.5%) and second (8.0%) spray at 14 days followed by Propiconazole 25% EC @ 500 ml/ha (PDI 4.7% and 8.3%), Azoxystrobin 23% SC @ 500 ml/ha (PDI 4.8% and 8.5 %). However, Picoxystrobin 22.52% SC @ 400 ml/ha showed PDI 5.3% and 9.0 % and PDC 68.2 % which was on par with Pyraclostrobin 20% WG @ 500 ml/ha (PDI 5.7% and 9.4 %) and PDC (66.3%) after 14 days of each spray, while, Hexaconazole 5% EC @ 500 ml/ha (PDI 6.3% and 10.1%) and PDC (63.6%) after 14 days of each spray and Mancozeb 75% WP @ 1500 gm/ha showed (6.4% and 12.6%) and PDC (57.4%) 14 days after first and second spray. Whereas, mean per cent disease intensity of control was recorded (19.48%) during study period. In all the treatments was significantly produced lower per cent disease intensity as compared to control. The highest per cent reduction over control i.e. 72.3% was recorded in i.e. Dinfenconazole 5% EC @ 250 ml/ha followed by spraying of Propiconazole 25% EC @ 500 ml/ha (71.3%) and Azoxystrobin 23% SC @ 500 ml/ha 70.2 % at 28 days of spray (Table 4.6) and (Fig.- 4.6a and 4.6b).









































































































